

## METHOD DEVELOPMENT AND VALIDATION OF METFORMIN BY USING REVERSE- PHASE HIGH PERFORMANCE LIQUID CHROMATOGRAPHY

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### ABSTRACT

The method development and validation of Metformin based on Ultra-Violet spectrophotometry. This compound belongs to the class of organic compounds known as biguanides. These are organic compound containing two N-linked guanidines. Metformin is an oral anti diabetic drug in preventing complications of type 2 diabetes and it is a good first line therapy for a over obese with type 2 diabetes, it is currently available in more than 60countries worldwide. As a result of the importance of this oral hypoglycaemic agent in the treatment of non insulin dependent diabetes mellitus. The reported methods include the chromatographic methods such as HPLC. HPLC has been reported for the determination of Metformin chromatographic methods, these are

complex, tedious, time consuming and are not suitable for routine analysis. However, Ultraviolet spectrophotometric methods which are simple and fast for ideal and routine analysis. We observed the maximum absorbance at 235&274nm for Metformin solution ELICO SL-159 make Ultra-Violet visible spectrophotometry spectrum model UV-2450, has been recorded when scanned in the ultraviolet region, using mobile aceto nitrile: phosphate buffer (30:70) with a flow rate of 1.0ml/minute and wave length at 225 nm. Thus, it was selected as the analytical wavelength. The development method was found to be simple, sensitive, accurate, precise and specific as evidenced by non-interference from the excipients presented in the marketed injection.

**KEYWORDS:** Metformin, reverse phase HPLC, ultraviolet spectroscopy, sensitive, accurate.

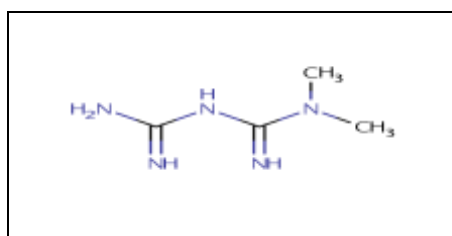
## INTRODUCTION

### Metformin

The discovery of metformin was began with synthesis of galegine like compounds derived from *Gallega officinalis*, it is a plant traditionally employed in Europe as a drug for diabetes treatment for many years. In 1950, Stern *et al.* discovered the clinical usefulness of metformin while working in Paris.<sup>[1]</sup> They observed that the dose-response of metformin was related to its glucose lowering capacity.

Metformin as a non-insuline oral hypoglycaemic which acts primarily in the liver by reducing glucose output and also by avoiding glucose uptake in the peripheral tissues. These effects are mediated by the activation of an upstream kinase, liver kinase B1 (LKB-1), which in turn regulates the downstream kinase adenosine monophosphate protein kinase (AMPK). AMPK phosphorylates a transcriptional co-activator, transducer of regulated CREB protein 2 (TORC2), resulting in its inactivation which consequently downregulates transcriptional events that promote synthesis of gluconeogenic enzymes and by the inhibition of mitochondrial respiration has also been proposed to contribute to the reduction of gluconeogenesis, as it reduces the energy supply to the cells for this process.<sup>[2]</sup> HPLC has been reported for the determination of Metformin chromatographic methods, these are complex, tedious, time consuming and are not suitable for routine analysis.

This compound belongs to the class of organic compounds known as biguanides.<sup>[3]</sup> These are organic compound containing two N-linked guanidines.



**Figure1. Structure of Metformin**

IUPAC Name: 1-carbamimidamido-N, N-dimethylmethanimidamide

Chemical formula: C<sub>4</sub>H<sub>11</sub>N<sub>5</sub>

Categories: Anti diabetic Agent

Weight: 129.1636

Description: White Crystalline Powder

Half life: 6.2 hours. Duration of action is 8-12 hours

**MATERIALS AND METHODS****Table 1: List of Instruments:**

S. No.	Instruments
1.	HITACHI L2130 with D Elite 2000 Software with Isocratic with UV-Visible Detector (L-2400),
2	ELICO SL-159 UV – Vis spectrophotometer
3	Electronic Balance (SHIMADZU ATY224)
4	Ultra Sonicator (Wensar wuc-2L)
5	Thermal Oven
6	Waters ODS (C <sub>18</sub> ) RP Column, 250 mm x 4.6 mm.
7	P <sup>H</sup> Analyzer (ELICO)
8	Triple Quartz Distillation Unit (BOROSIL)

**Table 2: List of Chemicals, Reagents and Standards Used:**

S.No	Name	Specifications		Manufacturer/Supplier
		Purity	Grade	
1.	Doubled distilled water	----	----	SD fine-Chem Ltd; Mumbai
2.	Methanol	99.9%	A.R.	Loba Chem; Mumbai.
3.	Acetonitrile	99.9%	HPLC	Loba Chem; Mumbai.
4.	Potassium dihydrogen orthophosphate	99.9	L.R.	SD fine-Chem Ltd; Mumbai
5.	glacial acetic acid	99.9	L.R.	SD fine-Chem Ltd; Mumbai
6.	Triethylamine	99.9%	L.R	SD fine-Chem Ltd; Mumbai

(L.R-Laboratory Reagent, A.R- Analytical Reagent, HPLC- High Performance Liquid Chromatography).

**Table 3: Solubility Study**

REAGENTS	SOLUBILITY
Methanol	Sparingly soluble
Ethanol	Slightly soluble
Acetonitrile	Soluble
1 M HCl	Insoluble
1M NaOH	Insoluble
Water	Insoluble

**METHOD DEVELOPMENT****HPLC Instrumentation & Conditions**

The HPLC system employed was HITACHI L2130 with D Elite 2000 Software with Isocratic with UV-Visible Detector (L-2400).

Standard & sample preparation for UV-spectrophotometer analysis<sup>[4]</sup>: 25 mg of Metformin standard was transferred into 25 ml volumetric flask, dissolved & make up to volume with mobile phase. Further dilution was done by transferring 0.5 ml of the above solution into a 10ml volumetric flask and make up to volume with mobile phase.

The standard & sample stock solutions<sup>[5]</sup> were prepared separately by dissolving standard & sample in a solvent in mobile phase diluting with the same solvent (After optimization of all conditions) for UV analysis. It scanned in the UV spectrum in the range of 200 to 400nm. This has been performed to know the maxima of Metformin, so that the same wave number can be utilized in HPLC UV detector for estimating the Metformin. While scanning the Metformin solution we observed the maxima at 265 nm. The UV spectrum has been recorded on ELICO SL-159 make UV – Vis spectrophotometer model UV-2450.

### Optimized Chromatographic Conditions

Column	: C <sub>18</sub> Develosil ODS HG-5 RP 150mm x 4.6mm 5µm particle size
Mobile Phase	: ACN: phosphate buffer = 30:70
Flow Rate	: 1.0ml/minute
Wave length	: 225 nm
Injection volume	: 20 µl
Run time	: 10 minutes
Column temperature	: Ambient
Sampler cooler	: Ambient

**Mobile Phase Preparation<sup>[6]</sup>**: The mobile phase used in this analysis consists of a mixture of phosphate buffer and Acetonitrile in a ratio of 70:30.700 ml of buffer was added and properly mixed with 300 ml of Acetonitrile and a homogenous solution is achieved. This mobile phase was filled and Sonicated for 15 minutes before the experiment.

**Sample & Standard Preparation for the Analysis:** 25 mg of Metformin standard was transferred into 25 ml volumetric flask, dissolved & make up to volume with mobile phase. Further dilution was done by transferring 0.5 ml of the above solution into a 10 ml volumetric flask and make up to volume with mobile phase. The sample was analysed by HPLC by using the above method and a very nicely resolved peak has been obtained at a Retention Time of about 2.80 min. The respective chromatogram is attached below.

## RESULT AND DISCUSSION

### Method development and optimization

The amount of drugs in Panex tablet was found to be 49.10 ( $\pm 0.498$ ) mg/tab for Metformin and 99.55 ( $\pm 0.494$ ) mg/tab for Metformin. To develop a precise, linear, specific & suitable stability indicating RP-HPLC method for analysis of Metformin, different chromatographic conditions were applied & the results observed are presented in previous chapters. Isocratic elution is simple, requires only one pump & flat baseline separation for easy and reproducible results. So, it was preferred for the current study over gradient elution. In case of RP-HPLC various columns are available, but here develosil ODS HG-5 RP C<sub>18</sub>, 5 $\mu$ m and 15cmx4.6mm internal diameter. Column was preferred because using this column peak shape, resolution and absorbance were good. Mobile phase & diluents for preparation of various samples were finalized after studying the solubility of API in different solvents of our disposal (methanol, acetonitrile, dichloromethane, water, 0.1N NaOH, 0.1NHCl). The drug was found to be highly soluble in methanol & dichloromethane and partially soluble in acetonitrile. Drug was insoluble in water.

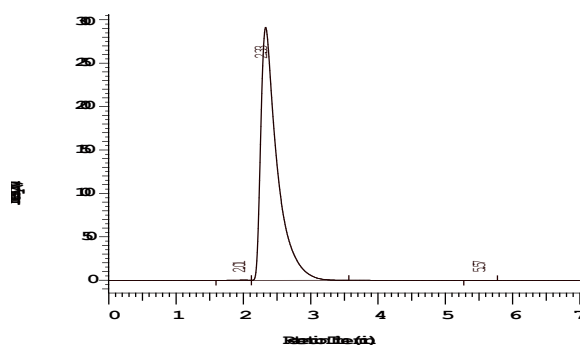


Figure.2. Chromatogram for Metformin (Rt2.39)

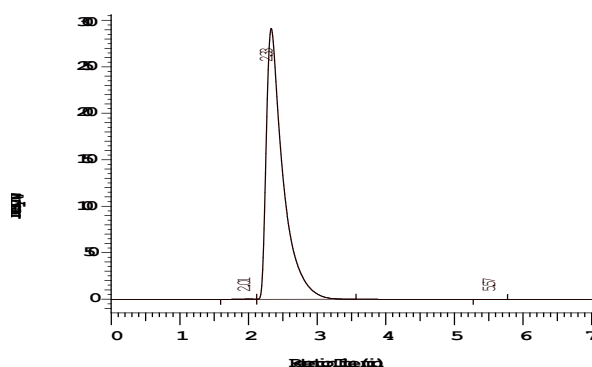


Figure 3. HPLC spectrum of Metformin (50 ppm) in optimized conditions (RT 2.33 min).

Using these solvents with appropriate composition newer methods can be developed and validated. Detection wavelength was selected after scanning the standard solution of drug over 200 to 400nm. From the U.V spectrum of Metformin it is evident that most of the HPLC work can be accomplished in the wavelength range of 240-300 nm conveniently. Further, a flow rate of 1 ml/min & an injection volume of 20  $\mu$ l were found to be the best analysis.

The result shows the developed method is yet another suitable method for assay which can help in the analysis of Metformin in different formulations.

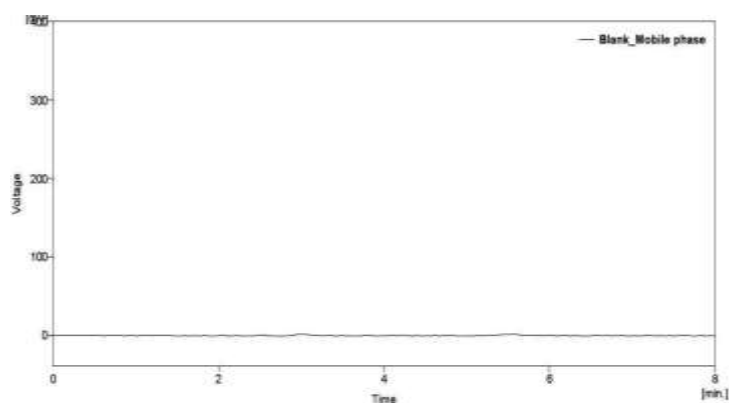


Figure. 4: Chromatogram for blank interference

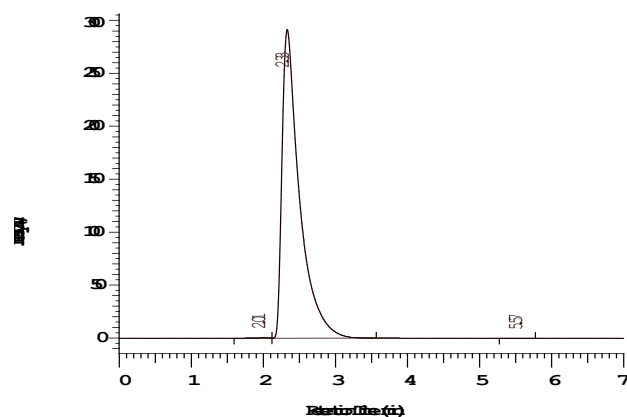


Figure. 5: Chromatogram for standard

#### METHOD VALIDATION

**Linearity & Range**<sup>[7]</sup>: The calibration curve showed good linearity in the range of 0-100  $\mu$ g/ml, for Metformin (API) with correlation coefficient ( $r^2$ ) of 0.998. A typical calibration curve has the regression equation of  $y = 69655x + 12056$  for Metformin.

Table. 4: Linearity Data

CONC.	AUC (n=6)
0	0
10	829757
20	1496593
30	2286179
40	2886713
50	3766816
100	6991760

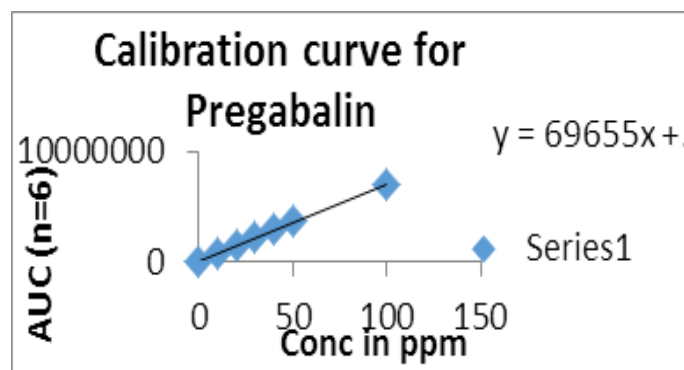


Figure 6: Linearity data of Metformin

**LOD & LOQ:** The Minimum concentration level at which the analyte can be reliably detected (LOD) & quantified (LOQ) were found to be 0.01 & 0.03  $\mu\text{g/ml}$  respectively.

#### Accuracy

**Recovery study<sup>[8]</sup>:** To determine the accuracy of the proposed method, recovery studies were carried out by adding different amounts (80%, 100% and 120%) of pure drug of Metformin were taken and added to the pre-analyzed formulation of concentration 10 $\mu\text{g/ml}$ . From that percentage recovery values were calculated.

Table. 5: Accuracy Readings:

Sample ID	Concentration ( $\mu\text{g/ml}$ )		%Recovery of Pure drug	Statistical Analysis
	Pure drug	Formulation		
S <sub>1</sub> : 80 %	16	20	101.3	Mean= 100.2733%
S <sub>2</sub> : 80 %	16	20	99.25	S.D. = 1.025004
S <sub>3</sub> : 80 %	16	20	100.27	% R.S.D.= 1.02221
S <sub>4</sub> : 100 %	20	20	99.14	Mean= 99.18%
S <sub>5</sub> : 100 %	20	20	99.29	S.D. = 0.096437
S <sub>6</sub> : 100 %	20	20	99.11	% R.S.D.= 0.097234
S <sub>7</sub> : 120 %	24	20	99.21	Mean= 99.46%
S <sub>8</sub> : 120 %	24	20	99.54	S.D. = 0.221133
S <sub>9</sub> : 120 %	24	20	99.63	% R.S.D. = 0.222334

### Precision

**Repeatability**<sup>[9]</sup>: The precision of each method was ascertained separately from the peak areas & retention times obtained by actual determination of five replicates of a fixed amount of drug Metformin (API). The percent relative standard deviations were calculated for metformin are presented in the.

**Table. 6: Relative standard deviations**

HPLC Injection, Replicates of Metformin	Retention Time	Area
Replicate – 1	2.33	1414087
Replicate – 2	2.33	1467177
Replicate – 3	2.33	1460891
Replicate – 4	2.33	1471809
Replicate – 5	2.33	1482648
Replicate – 6	2.33	1489902
Average	2.33	1464419
Standard Deviation	0.0	26794.24
% RSD	0.0	1.829684

### Intra-day & inter-day precision<sup>[10]</sup>

The intra & inter day variation of the method was carried out & the high values of mean assay & low values of standard deviation & % RSD (% RSD < 2%) within a day & day to day variations for Metformin revealed that the proposed method is precise.

**Table.7: Results of intra-day & inter-day precision:**

Conc. of Metformin (API) (µg/ml)	Observed Conc. Of Metformin (µg/ml) by the proposed method			
	Intra-Day		Inter-Day	
	Mean (n=6)	% RSD	Mean (n=6)	% RSD
18	18.03	1.03	18.01	0.46
20	20.09	0.51	19.94	0.28
22	22.14	0.19	22.19	0.15

### System Suitability Parameter<sup>[11]</sup>

System suitability testing is an integral part of many analytical procedures. The tests are based on the concept that the equipment, electronics, analytical operations and samples to be analyzed constitute an integral system that can be evaluated as such. Following system suitability test parameters were established. The data are shown in below table:

**Table. 8: Data of System Suitability Parameter**

S.No.	Parameter	Limit	Result
1	Resolution	$R_s > 2$	9.15
2	Asymmetry	$T \leq 2$	Metformin =0.12
3	Theoretical plate	$N > 2000$	Metformin =3246



**Method Robustness<sup>[12]</sup>**

Influence of small changes in chromatographic conditions such as change in flow rate ( $\pm 0.1$  ml/min), Temperature ( $\pm 2^{\circ}\text{C}$ ), Wavelength of detection ( $\pm 2$  nm) & acetonitrile content in mobile phase ( $\pm 2\%$ ) studied to determine the robustness of the method are also in favour of (Table-4, % RSD < 2%) the developed RP-HPLC method for the analysis of Metformin(API).

**Table. 9: Result of method robustness test**

Change in parameter	% RSD
Flow (1.1 ml/min)	0.04
Flow (0.9 ml/min)	0.09
Temperature ( $27^{\circ}\text{C}$ )	0.01
Temperature ( $23^{\circ}\text{C}$ )	0.13
Wavelength of Detection (227 nm)	0.07
Wavelength of detection (223 nm)	0.05

**Assay of Metformin in Dosage Form****Estimation of Metformin in tablet Dosage Form****Metformin 20 mg**

Twenty tablets were taken and the I.P. method was followed to determine the average weight. Above weighed tablets were finely powdered and triturated well. A quantity of powder equivalent to 100 mg of drug was transferred to 100 ml volumetric flask and 70 ml of HPLC grade methanol was added and solution was sonicated for 15 minutes, there after volume was made up to 100 ml with same solvent. Then 10 ml of the above solution was diluted to 100 ml with HPLC grade methanol.<sup>[13]</sup> The solution was filtered through a membrane filter (0.45  $\mu\text{m}$ ) and sonicated to degas. From this stock solution (3.5 ml) was transferred to five different 10 ml volumetric flasks and volume was made up to 10 ml with same solvent system. The solution prepared was injected in five replicates into the HPLC system and the observations were recorded. A duplicate injection of the standard solution was also injected into the HPLC system and the peak areas were recorded. The data are shown below,

**ASSAY:**

$$\text{Assay \%} = \frac{AT}{AS} \times \frac{WS}{DS} \times \frac{DT}{WT} \times \frac{P}{100} \times \text{Avg. Wt} = \text{mg/tab}$$

Where: AT = Peak Area of test obtained with test preparation

AS = Peak Area of standard obtained with standard preparation

WS = Weight of working standard taken in mg

WT = Weight of sample taken in mg

DS = Dilution of Standard solution

DT = Dilution of sample solution

P = Percentage purity of working standard

**Table. 10: Estimation of Metformin in Capsule Dosage Form**

Brand name of tablet	Labelled amount of Drug (mg)	Mean ( $\pm$ SD) amount (mg) found by the proposed method (n=6)	Assay + % RSD
Panex (Synapse (Micro Labs Ltd))	20	19.91 ( $\pm$ 0.498)	99.55 (0.494)

## CONCLUSION

A sensitive & selective RP-HPLC method has been developed & validated for the analysis of Metformin Active Pharmaceutical Ingredient. Further the proposed RP-HPLC method has excellent sensitivity, precision and reproducibility. Above results shows the developed RP-HPLC method is yet another suitable method for assay and purity which can help in the analysis of Metformin in different formulations.

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