

# Comparative clinical evaluation of *Boerhavia diffusa* root extract with standard Enalapril treatment in Canine chronic renal failure

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## ABSTRACT

**Background:** Complementing herbal drugs with conservative modern treatment could improve renal condition in canine chronic renal failure (CRF). **Objective:** In this study, clinical evaluation of *Boerhavia diffusa* root extract was carried out in CRF in dogs in comparison with standard enalapril. **Materials and Methods:** A total of 20 dogs of mixed breeds suffering from CRF from 1 to 2 months were divided into two groups ( $n = 10$ ) and treated as follows: Group I - Enalapril at 0.5 mg/kg p.o. once daily for 90 days + amoxicillin and cloxacillin at 25 mg/kg i.m. once daily for 1-week; Group II - *B. diffusa* root extract at 500 mg p.o per dog daily for 90 days. Both groups were maintained on a supportive fluid therapy. The data were analyzed using paired *t*-test and one-way ANOVA followed by Dunnett's *post-hoc* test. **Results:** CRF caused a significant ( $P < 0.05$ ) increase in systolic and diastolic blood pressure, serum creatinine, urea nitrogen, sodium, potassium, phosphorus, urinary protein, alkaline phosphatase (ALP), and glutamyl transferase (GGT). A significant ( $P < 0.05$ ) decrease in hemoglobin and total erythrocyte count (TEC) was also observed. Nephrosonography revealed indistinct corticomedullary junction, altered renal architecture, hyper-echoic cortex, medulla, and sunken kidneys. Both the treatments significantly ( $P < 0.05$ ) reduced systolic and diastolic blood pressure by day 30. Serum Creatinine, urea nitrogen, phosphorus, urinary protein, ALP, and GGT showed significant ( $P < 0.05$ ) reduction by day 60 in both the treatments. However, potassium levels were normalized only by *B. diffusa* root extract treatment by day 30. Both the treatments failed to show a significant improvement in nephrosonographic picture even after 90 days posttreatment. **Conclusion:** In conclusion, the efficacy of *B. diffusa* root extract was comparable to standard enalapril treatment of CRF in dogs.

**Key words:** *Boerhavia diffusa*, chronic renal failure, dogs, enalapril, nephrosonography

## INTRODUCTION

Chronic renal failure (CRF) or chronic kidney disease is a common kidney disease in dogs with a prevalence of 0.05–3.74%. The risk factors for CRF include old age, specific breeds, smaller body size, periodontal

disease, and obesity.<sup>[1]</sup> Standard therapy for CRF is aimed at the management of proteinuria, inhibition of renin-angiotensin-aldosterone system, correcting fluid balance, and hypertension.<sup>[2]</sup> But with the progression of CRF to end-stage disease, renal function can be regenerated only by kidney transplantation or dialysis, which is costlier and unaffordable in veterinary cases. Hence, Ayurvedic drugs can be used to complement modern medicines to reverse kidney damage in animals.<sup>[3]</sup>

Herbs are increasingly becoming popular for the treatment of various diseases in both human and veterinary practice. Several instances of medicinal properties of plants and plant products are well-documented in animal models such as anti-hyperlipidemic activity,<sup>[4]</sup> anti-diabetic activity,<sup>[5]</sup> protective activity against toxicities produced by mycotoxins,<sup>[6]</sup> pesticides,<sup>[7]</sup> and heavy metals.<sup>[8]</sup> Further, several plant products are found to be safe through safety assessment as per OECD guidelines.<sup>[9–11]</sup> Recently, phytochemicals are being used for the synthesis of nanoparticles, which are effective and safe in several diseases.<sup>[12,13]</sup>

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Received: 22-Dec-2014

Revised: 04-Jun-2015

Accepted: 06-Jun-2015

### Access this article online

Quick Response Code:



Website:

www.jaim.in

DOI:

10.4103/0975-9476.166390

*Boerhavia diffusa* (Family: Nyctaginaceae) is commonly known as Raktapunarnava, Shothaghni, Kathillaka, Kshudra, Varshabhu, Raktapushpa, Varshaketu, and Shilatika.<sup>[14,15]</sup> The plant is also called “Punarnava,” due to its ability to regenerate in rainy season with the help of perennial roots after the aerial parts get dried up completely in summer.<sup>[16]</sup> In Ayurveda, the plant is considered to be light (Laghu), dry (Ruksha) and hot potency (Ushna veerya) and the properties like: Rasa-Madhura, Tikta, Kashaya; Veerya-Ushana; Vipaka-Madhura and Karma-Anulomana, Shothahara and is considered to alleviate all three doshas.<sup>[17]</sup>

The roots of *B. diffusa* contain many rotenoids.<sup>[18-21]</sup> Further, it also has Punarnavoside, a phenolic glycoside,<sup>[22-23]</sup> C-methyl flavone<sup>[24]</sup> and 6.0% potassium nitrate, and ursolic acids.<sup>[25]</sup> *B. diffusa* was reported to offer significant protection against kidney disease<sup>[26]</sup> and urolithiasis.<sup>[27]</sup> The regenerative effects of *B. diffusa* on kidneys is also reported.<sup>[28]</sup> However, the therapeutic efficacy of root extract of *B. diffusa* for the treatment of CRF is not extensively studied in veterinary cases. Hence, this study was aimed at investigating nephroprotective effect of *B. diffusa* for the treatment of CRF in dogs in comparison with modern conservative treatment.

## MATERIALS AND METHODS

### Chemicals used

Boerhavia root hydro-alcoholic extract (Himalaya Punarnava-Himalaya Drug Company, India; containing 250 mg hydro-alcoholic extract per capsule), enalapril (Canvas 5 mg-Zyudus Cadila, India); ampicillin and cloxacillin (Novaclox 1 g-Cipla, India), metaclopramide (Perinorm 5 mg/mL-IPCA Laboratories Ltd., India); ranitidine (Aciloc 50 mg/mL-Cadila Pharmaceuticals, India), Ringer’s lactate (Basol Infusion-Cadila Pharmaceuticals, India); B-complex (Polybion-Merck, India) were used in the study.

### Animals used

Dogs belonging to the breeds Spitz, German Shepard, Labrador Retriever, Great Dane, Doberman pinscher and mongrels of both sexes aged between 8 and 12 years of age suffering from renal failure were included in the study. Healthy dogs of Animal Care Land, Tirupati were used as controls.

### Clinical cases

Dogs presented to the Teaching Veterinary Clinical Complex of the College with clinical and nephrosonographic changes suggestive of CRF, serum creatinine between 3.0 and 5.0 mg/dL and without anemia or ascites from 1 to 2 months were included in the study. Institutional Animal Ethic Committee approval was obtained prior to the start

of the study. A total of 20 dogs with the above criteria were randomly divided into two treatment groups. Group I dogs were treated with enalapril at 0.5 mg/kg p.o once daily for 90 days + amoxicillin and cloxacillin at 25 mg/kg i.m once daily for 1-week; Group II dogs were treated with *B. diffusa* root extract at 500 mg per animal p.o, once daily for 90 days. Both the groups were maintained on a supportive therapy consisting of ringer’s lactate infusion at 30 mL/kg i.v., once daily for correcting electrolyte imbalance; metoclopramide at 0.2 mg/kg i.m., once daily for preventing uremia-induced-vomition; ranitidine at 2 mg/kg i.m., once daily as H<sub>2</sub>-antagonist for decreasing gastric acid production and B-complex at 1 mL/dog i.m., once daily for improving status of water-soluble B-vitamins. The owners were advised to provide low salt and low protein diet and to increase energy density of the feed. Both the treatment groups were compared with 10 apparently healthy dogs of different breeds aged 3–5 years.

Detailed history, clinical observations, blood pressure monitoring, serum biochemical profile, urinalysis, and nephrosonography were carried out at monthly interval up to 3 months.

### Blood pressure monitoring

For measuring blood pressure, human wrist model automatic oscillometric sphygmomanometer (BPL Ltd., India) was used. The dog was positioned in sternal recumbency, and the cuff was placed on the left forelimb region. The transducer was positioned on the medial aspect of the arm over the median artery, and the Velcro was wrapped around the foreleg [Figure 1]. The average of five consecutive readings was taken as the blood pressure.

### Sero-biochemical profile

Serum was obtained from 3 mL of blood collected from saphenous vein and parameters such as creatinine, urea



**Figure 1:** Placement of sphygmomanometer on the left forelimb of dog in sternal recumbency

nitrogen, total protein, sodium, potassium, calcium, phosphorus were analyzed using standard kits supplied by span diagnostics Ltd., Surat using star 21 semi-auto biochemistry analyzer (Rapid Diagnostic Pvt., Ltd., Delhi)

### Urinalysis

Five milliliters of mid-stream or cystecentesized urine was collected and urine pH, specific gravity, and protein were determined using URISCAN dip sticks. Later, the urine was centrifuged at 1500 rpm for 5 min, and the sediment was examined for casts, pus cells, and other sediments. Alkaline phosphatase (ALP) and gamma glutamyl transferase (GGT) were estimated from the supernatant urine using standard kits supplied by Accurex Biomedical Pvt., Ltd, Mumbai.

### Nephrosonography

The hair on the abdomen was shaved midway up to the body wall over the right and left caudal intercostal spaces. Nephrosonography was performed in either dorsal or sternal recumbency using IXOS vet-ultrasound machine supplied by Esoate Pie Medical, Netherlands. A linear array of 3.5, 5.0, and 7.5 mHz probes were used for small, medium, or large dogs, respectively. The left kidney was imaged caudal to the greater curvature of the stomach, caudo-dorsal to the spleen, later to the aorta, and left adrenal gland at the level of L2– L4 vertebrae. The right kidney was imaged caudal to right liver lobes, lateral to the caudal vena cava and right adrenal gland at the level of L1–L3 vertebrae.<sup>[29]</sup>

Sonograms were evaluated for information on renal architecture specifically including focal, multifocal of diffuse alterations in renal cortical, medullary, sinusal, and peripheral echogenicity. In addition, cortical and medullary echogenicity were compared subjectively with hepatic and splenic parenchymal echogenicity. The echogenicity of the identifiable lesion, as seen on the gray scale two-dimensional images were classified subjectively as normal, increased (hyperechoic), decreased (hypoechoic), and absent compared to normal echo pattern for canine kidney.<sup>[30]</sup>

### Statistical analysis

The data for various parameters were expressed as mean ± standard error. In both the groups, after treatment values at different time intervals (30, 60, and 90 days) were

compared with before treatment values (0 day) using paired *t*-test. Similarly, the control values were compared with different time periods (0, 30, 60, and 90 days) using one-way ANOVA followed by Dunnett's *post-hoc* test using Statistical package for social sciences (SPSS) 19.0V (IBM SPSS, v 19.0, Armonk, NY). The level of significance was set at  $P < 0.05$ .

## RESULTS

The predominant symptoms in CRF dogs were anorexia, vomiting, dullness, weight loss, oral ulcers and in few cases polydipsia, pale mucosa, recumbency, and blindness were also observed before treatment. A significant ( $P < 0.05$ ) increase in both systolic and diastolic arterial pressure was observed in CRF affected dogs compared to control. Treatments with enalapril in Group I and *B. diffusa* root extract in Group II significantly ( $P < 0.05$ ) reduced both systolic and diastolic blood pressure by day 30 and were comparable to control dogs [Table 1].

The hemoglobin (Hb) and total erythrocyte concentrations (TEC) in CRF dogs were significantly ( $P < 0.05$ ) decreased compared to control dogs. Both enalapril and *B. diffusa* root extract treatment could significantly ( $P < 0.05$ ) increase Hb levels by day 60; however, only in *B. diffusa* root extract treatment, the Hb values were comparable to normal by day 90. Both treatments failed to show any significant improvement in TEC even after 90 days posttreatment [Table 2].

In CRF affected dogs, a significant ( $P < 0.05$ ) elevation of serum creatinine, urea nitrogen, total protein, albumin, and phosphorus levels compared to control group before treatment. The urine of CRF dogs revealed casts, epithelial cells, red blood cells in the sediment. A significant ( $P < 0.05$ ) decrease in specific gravity and a significant ( $P < 0.05$ ) increase in urinary protein [Figure 2], ALP, and GGT were also elevated compared to control dogs on day 0. Both the treatments significantly ( $P < 0.05$ ) decreased serum creatinine, urea nitrogen, urinary protein [Figure 2], and urine ALP and GGT by day 60. However, potassium [Figure 3] and phosphorus levels showed significant ( $P < 0.05$ ) reduction by day 30 only in *B. diffusa* root extract treatment. Enalapril treatment could significantly ( $P < 0.05$ ) reduce only phosphorus level by

**Table 1: Mean blood pressure in treatment groups at various time intervals**

Parameter	Control	Treatment	o day	30 days	60 days	90 days <sup>†</sup>
Systolic pressure (mmHg)	121.1±1.80	I	131.20±4.62 <sup>#</sup>	123.30±1.63 <sup>**</sup>	120.57±3.27 <sup>**</sup>	117.40±1.03 <sup>**</sup>
		II	136.8±1.76 <sup>#</sup>	121.60±1.44 <sup>**</sup>	121.75±1.28 <sup>**</sup>	114.29±2.85 <sup>**</sup>
Diastolic arterial pressure (mmHg)	71.1±1.33	I	83.70±2.75 <sup>#</sup>	70.00±3.19 <sup>**</sup>	71.70±4.30 <sup>*</sup>	71.40±2.54 <sup>*</sup>
		II	79.4±2.56 <sup>#</sup>	72.80±1.98 <sup>**</sup>	74.25±1.93 <sup>*</sup>	68.00±1.92 <sup>**</sup>

Values are mean±SE (n=10); one-way ANOVA followed by Dunnett's *post-hoc* test for comparing control with treatments at different time periods; paired *t*-test for comparing o day with other time periods using SPSS 19.0 V software. <sup>#</sup>Significant difference with control group, <sup>\*</sup>Significant difference with o day, <sup>\*\*</sup> $P < 0.05$ , <sup>\*\*\*</sup> $P < 0.01$ , <sup>†</sup>Due to mortality, 90 days mean was computed with five dogs in treatment I and eight dogs in treatment II. SE: Standard error



day 60 [Tables 3 and 4] but failed to improve potassium level.

Nephrosonography of normal dogs revealed that the renal cortical echogenicity of the left kidney was less than adjacent spleen; right kidney cortical echogenicity was less than the adjacent liver [Figure 4]. The medulla was hypoechoic, round with well-defined corticomedullary junction [Figure 5]. Pelvis was hyperechoic. In dogs affected with CRF, the cortex was hyperechoic with indistinct corticomedullary junction [Figure 6], altered renal architecture, and sunken kidneys [Figure 7]. However, both the treatments failed to show significant improvement in nephrosonogram even after 90 days of treatment.

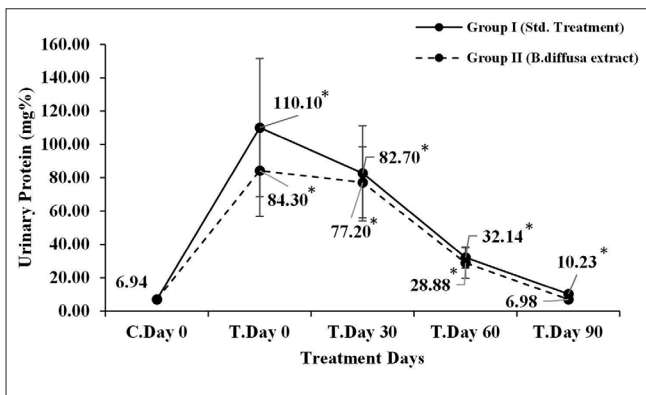
**DISCUSSION**

CRF is an important clinical condition in dogs which results from reduced renal function and to impaired homeostasis. As the clinical signs of CRF are nonspecific, many cases go unnoticed in veterinary practice. The treatment of CRF is also an economic constraint for the owner. The predominant clinical signs in CRF dogs were anorexia, vomiting, dullness, weight loss, oral ulcers, polyuria, polydipsia, pallor of mucous membrane, melena,

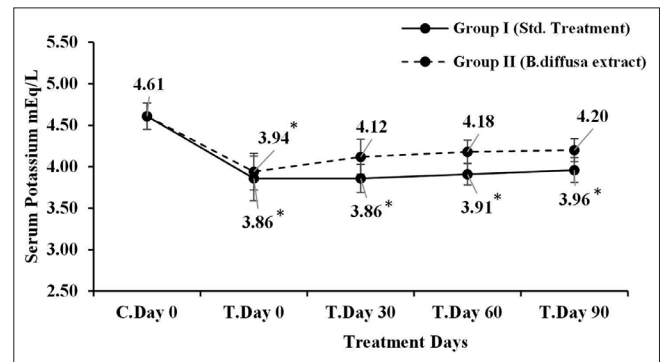
recumbency, and blindness. These signs are consistent with earlier findings.<sup>[31-33]</sup> Renal dysfunction leads to uremia, which stimulates chemoreceptor trigger zone, resulting in anorexia and vomition.<sup>[34]</sup> Weight loss and dullness are directly linked to inadequate calorie intake, catabolic effects of uremia, and intestinal malabsorption secondary to uremic gastroenteritis.<sup>[35]</sup> Pallor mucous membrane due to anemia, a characteristic symptom of advanced CRF, results from decreased erythropoietin production by damaged kidneys.<sup>[36]</sup>

In this study, the clinical cases showed the signs of improvement between 15 and 30 days of treatment in both the groups. Conservative therapy of CRF dogs consisted of symptomatic and supportive therapy designed to correct the deficiencies and excess in fluids, electrolytes, acid-base, and nutritional imbalances and thereby minimizing the clinical and pathological consequences of reduced renal function.<sup>[2]</sup> After 90 days of treatment, moderate improvement in appetite, body weight gain and improvement in behavior in survived dogs were noticed in both the groups. However, five dogs in enalapril treatment and two dogs in *B. diffusa* treatment died between 60 and 90 days posttreatment.

Systolic arterial and diastolic arterial pressure showed a significant ( $P < 0.05$ ) increase in CRF dogs



**Figure 2:** Urinary protein levels in control and treatment groups. *Boerhavia diffusa* extract could normalize urinary protein by day 90, whereas, no improvement was observed in conventional treatment. Std.: Standard, C: Control, T: Treatment. \*Significant ( $P < 0.05$ ) difference with control group



**Figure 3:** Serum potassium levels in control and treatment groups. *Boerhavia diffusa* could normalize serum potassium level by day 30, whereas, no improvement was observed in conventional treatment. Std.: Standard, C: Control, T: Treatment. \*Significant ( $P < 0.05$ ) difference with control group

**Table 2: Mean hematological parameters in treatment groups at various time intervals**

Parameter	Control	Treatment	o day	30 days	60 days	90 days <sup>†</sup>
Hb (g%)	15.28±0.39	I	11.94±0.53 <sup>#</sup>	11.92±0.48 <sup>#</sup>	13.30±0.22 <sup>#,*</sup>	13.79±0.35 <sup>#,**</sup>
		II	13.12±0.40 <sup>#</sup>	13.28±0.39 <sup>#</sup>	14.25±0.33 <sup>#,*</sup>	14.51±0.37 <sup>*</sup>
TEC (10 <sup>6</sup> /mm <sup>3</sup> )	7.58±0.32	I	4.26±0.31 <sup>#</sup>	4.29±0.27 <sup>#</sup>	3.58±0.18 <sup>#</sup>	4.69±0.13 <sup>#</sup>
		II	4.63±0.37 <sup>#</sup>	4.44±0.37 <sup>#</sup>	4.03±0.17 <sup>#</sup>	4.80±0.08 <sup>#</sup>
Total leucocyte count (10 <sup>3</sup> /mm <sup>3</sup> )	9.61±0.33	I	12.93±1.56	11.50±0.87	12.42±1.30	11.46±0.71
		II	10.06±1.13	10.72±1.02	13.43±1.83	1.24±0.88

Values are mean±SE (n=10); one-way ANOVA followed by Dunnett's *post-hoc* test for comparing control with treatments at different time periods; paired *t*-test for comparing o day with other time periods using SPSS 19.0 V software. <sup>#</sup>Significant difference with control group, <sup>\*</sup>Significant difference with o day, <sup>\*\*</sup> $P < 0.05$ , <sup>\*\*\*</sup> $P < 0.01$ , <sup>†</sup>Due to mortality, 90 days mean was computed with five dogs in treatment I and eight dogs in treatment II. SE: Standard error, Hb: Hemoglobin, TEC: Total erythrocyte count

**Table 3: Serum biochemical profile in treatment groups at various time intervals**

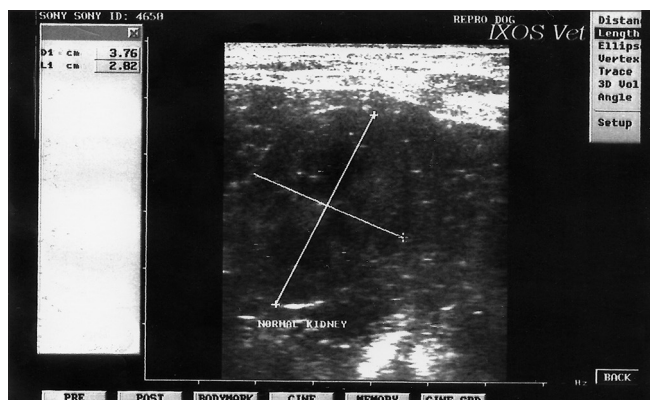
Parameters	Control	Treatment	o day	30 days	60 days	90 days†
Creatinine (mg%)	0.45±0.05	I	4.27±0.20 <sup>##</sup>	4.01±0.22 <sup>##</sup>	2.92±0.17 <sup>**##</sup>	2.41±0.09 <sup>**##</sup>
		II	4.19±0.18 <sup>##</sup>	4.03±0.20 <sup>##</sup>	3.11±0.14 <sup>**##</sup>	1.43±0.12 <sup>**##</sup>
Urea nitrogen (mg%)	19.00±2.24	I	162.70±20.02 <sup>##</sup>	148.80±10.73 <sup>##</sup>	138.00±10.25 <sup>**##</sup>	127.40±3.93 <sup>**##</sup>
		II	141.10±9.39 <sup>##</sup>	115.10±9.33 <sup>**##</sup>	74.00±5.79 <sup>**##</sup>	40.85±3.74 <sup>**##</sup>
Total protein (g%)	7.26±0.31	I	6.37±0.32	6.53±0.26	6.48±0.13	6.90±0.10
		II	6.55±0.17	6.65±0.13	6.69±0.16	6.90±0.14
Albumin (g%)	3.54±0.16	I	3.24±0.22	3.29±0.20	3.20±0.16	3.13±0.13
		II	3.80±0.24	3.65±0.24	3.62±0.14	3.64±0.32
Sodium (mEq/L)	145.22±4.19	I	161.80±4.76 <sup>##</sup>	159.60±4.10 <sup>##</sup>	153.63±2.18 <sup>**##</sup>	151.43±1.09 <sup>**##</sup>
		II	170.80±3.84 <sup>##</sup>	163.60±3.80 <sup>##</sup>	154.00±3.87 <sup>**#</sup>	150.80±2.38 <sup>**#</sup>
Potassium (mEq/L)	4.61±0.16	I	3.86±0.27 <sup>#</sup>	3.86±0.17 <sup>#</sup>	3.91±0.13 <sup>#</sup>	3.96±0.15 <sup>#</sup>
		II	3.94±0.22 <sup>#</sup>	4.12±0.21	4.18±0.14	4.20±0.14
Phosphorus (mg%)	3.88±0.23	I	6.26±0.38 <sup>##</sup>	5.24±0.48 <sup>#</sup>	4.37±0.45 <sup>**</sup>	3.99±0.45 <sup>**</sup>
		II	6.55±0.20 <sup>##</sup>	4.59±0.29 <sup>**</sup>	3.72±0.07 <sup>**</sup>	3.65±0.07 <sup>**</sup>
Calcium (mg%)	9.75±0.49	I	9.11±0.22	9.42±0.14	9.46±0.25	9.40±0.18
		II	9.40±0.36	9.08±0.15	9.34±0.14	9.93±0.09

Values are mean±SE (n=10); one-way ANOVA followed by Dunnett's *post-hoc* test for comparing control with treatments at different time periods; paired *t*-test for comparing o day with other time periods using SPSS 19.0 V software. <sup>#</sup>Significant difference with control group, <sup>\*</sup>Significant difference with o day, <sup>\*\*</sup>*P*<0.05, <sup>\*\*#</sup>*P*<0.01, <sup>†</sup>Due to mortality, 90 days mean was computed with five dogs in treatment I and eight dogs in treatment II. SE: Standard error

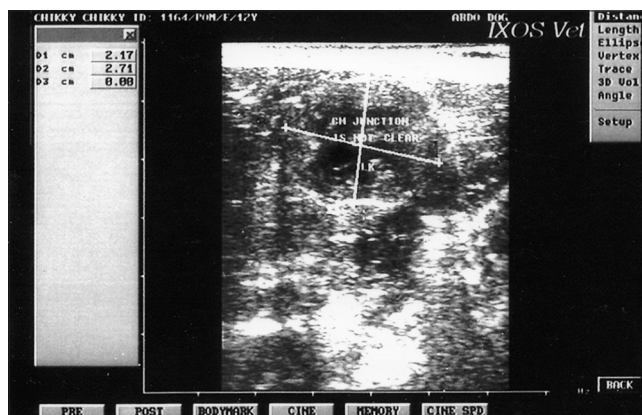
**Table 4: Urinalysis in treatment groups at various time intervals**

Parameters	Control	Treatment	o day	30 days	60 days	90 days†
pH	6.67±0.17	I	6.62±0.07	6.74±0.04	6.71±0.06	6.76±0.08
		II	6.75±0.10	6.76±0.06	6.76±0.06	6.77±0.10
Specific gravity	1.036±0.02	I	1.022±0.002	1.027±0.002	1.033±0.002	1.034±0.003
		II	1.020±0.002	1.023±0.002	1.029±0.002	1.035±0.003
Urinary protein (mg%)	6.94±0.55	I	110.10±41.54 <sup>##</sup>	82.70±28.57 <sup>##</sup>	32.14±6.38 <sup>**##</sup>	10.23±1.45 <sup>**#</sup>
		II	84.30±27.47 <sup>##</sup>	77.20±21.36 <sup>##</sup>	28.88±9.20 <sup>**#</sup>	6.98±1.22 <sup>**</sup>
Urinary ALP (mmol/L)	1.62±0.09	I	9.55±0.79 <sup>##</sup>	8.64±0.66 <sup>##</sup>	5.37±0.60 <sup>**##</sup>	2.19±0.51 <sup>**</sup>
		II	8.78±0.60 <sup>##</sup>	6.81±0.46 <sup>**##</sup>	4.32±0.59 <sup>**##</sup>	1.36±0.16 <sup>**</sup>
Urinary GGT (mmol/L)	1.55±0.09	I	8.11±0.46 <sup>##</sup>	7.43±0.52 <sup>##</sup>	4.93±0.54 <sup>**##</sup>	1.91±0.28 <sup>**</sup>
		II	7.69±0.40 <sup>##</sup>	5.68±0.39 <sup>**##</sup>	4.08±0.55 <sup>**##</sup>	1.24±0.12 <sup>**</sup>

Values are mean±SE (n=10); one-way ANOVA followed by Dunnett's *post-hoc* test for comparing control with treatments at different time periods; paired *t*-test for comparing o day with other time periods using SPSS 19.0 V software. <sup>#</sup>Significant difference with control group, <sup>\*</sup>Significant difference with o day, <sup>\*\*</sup>*P*<0.05, <sup>\*\*#</sup>*P*<0.01, <sup>†</sup>Due to mortality 90 days mean was computed with five dogs in treatment I and eight dogs in treatment II. SE: Standard error, ALP: Alkaline phosphatase, GGT: Gamma glutamyl transferase



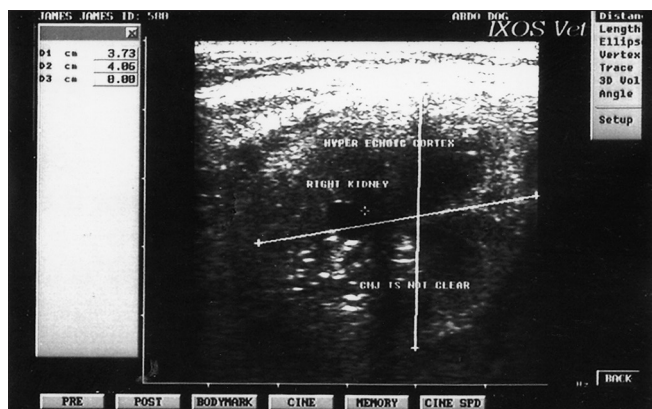
**Figure 4:** Nephrosonogram showing normal kidney in control dogs



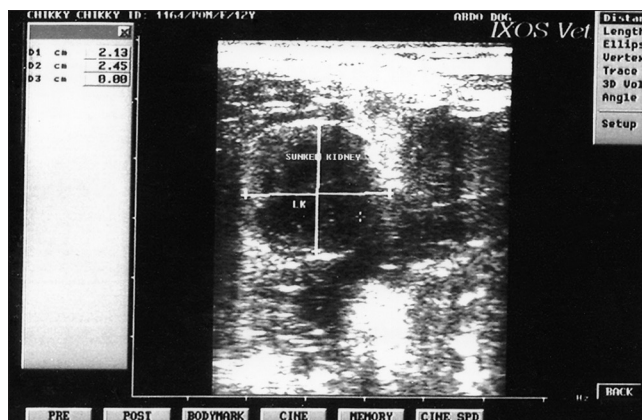
**Figure 5:** Nephrosonogram showing hyperechoic medulla without clear corticomedullary junction in dogs with chronic renal failure

compared to healthy dogs.<sup>[37-39]</sup> Both the treatments significantly (*P* < 0.05) decreased the systolic and diastolic arterial pressure by day 30. Enalapril, an angiotensin converting enzyme inhibitor, is reported to possess

anti-hypertensive activity and was earlier used successfully in several cases of CRF in dogs.<sup>[40-42]</sup> The anti-hypertensive activity of *B. diffusa* root extract can be attributed to



**Figure 6:** Nephrosonogram showing hyperechoic cortex without clear corticomedullary junction in dogs with chronic renal failure



**Figure 7:** Nephrosonogram showing shrunken kidney in dogs with chronic renal failure

punarnavoside component, which was reported to possess anti-hypertensive property.<sup>[43]</sup> The root extract of *B. diffusa* was successfully used by ayurvedic practitioners for management of CRF in human beings.<sup>[44,45]</sup>

In CRF, about 2/3 of the nephrons in kidney are damaged, which results in decreased water conservation and loss of several important substances.<sup>[46]</sup> In this study, the mean values of serum urea nitrogen, creatinine, sodium, and phosphorous were significantly ( $P < 0.05$ ) elevated in CRF dogs compared to control.<sup>[34,47-49]</sup> The raised serum urea nitrogen and creatinine levels in CRF dogs could be due to retention of nitrogenous substances<sup>[48,50]</sup> due to reduced glomerular filtration rate and decreased excretory rate of kidneys.<sup>[48,50,51]</sup> In addition, gastrointestinal hemorrhages also contribute to increased urea nitrogen due to enhanced absorption of nitrogenous compounds.<sup>[56]</sup> Enalapril treatment significantly ( $P < 0.05$ ) decreased urea nitrogen, creatinine, sodium, and phosphorus levels by day 60 compared to day 0. However, enalapril treatment failed to improve serum potassium level even after 90 days of treatment. This indicated stable renal function and delayed progression of the renal disease by enalapril treatment.<sup>[55]</sup> Similar reduction results were seen with *B. diffusa* root extract treatment except that earlier response (by day 30) in terms of significantly ( $P < 0.05$ ) decreased urea nitrogen and phosphorus was observed. Further, the decreased potassium levels were restored to normal by day 30 in *B. diffusa* root extract treatment, which can be attributed to the potassium nitrate content (6%) in *B. diffusa* root extract.<sup>[43]</sup> Similarly, the elevated sodium levels in CRF was significantly ( $P < 0.05$ ) decreased by day 60 in *B. diffusa* treatment consequent to improved renal function. *B. diffusa* has a diuretic effect similar to furosemide, a potent loop diuretic<sup>[26]</sup> and is responsible for the enhanced elimination of metabolic wastes.<sup>[43,57]</sup>

The total protein and albumin levels showed no significant change in CRF dogs compared to control dogs. This is possibly due to improved appetite and decreased catabolic

effects by virtue of the partial restoration of renal function and anti-proteinuric effect of enalapril<sup>[53-55]</sup> and *B. diffusa*.<sup>[43]</sup> However, earlier works<sup>[47]</sup> observed hypoproteinemia and hypoalbuminemia in CRF dogs and attributed the loss of albumin through glomeruli, owing to its small size, as the possible explanation.<sup>[52]</sup>

Elevated markers enzymes such as ALP and GGT are indicative of renal damage.<sup>[58,59]</sup> In CRF, as a consequence of kidney damage, the concentrating ability of the kidney is lost leading to polyuria and decreased specific gravity of urine.<sup>[34]</sup> Similarly, glomerular damage results in increased urinary protein excretion.<sup>[50]</sup> The reduction in urinary protein excretion could be attributed to the anti-proteinuric effect of enalapril<sup>[54,55]</sup> and diuretic action of *B. diffusa* in treatment Group II.<sup>[26,60]</sup>

The ultrasonographic changes in CRF revealed hyperechoic cortex, indistinct corticomedullary junction, and hyperechoic medulla. Several authors reported overall increase in echogenicity (hyperechoic) and reduced corticomedullary definitions in dogs with chronic inflammatory and end-stage renal diseases.<sup>[30,61-63]</sup> The deposition of calcium in renal cortex is possible the explanation for increased echogenicity.<sup>[61]</sup>

CRF is a serious progressive and irreversible disease usually seen in older dogs with poor prognosis. Between day 60 and 90, five dogs in enalapril group and two dogs in *B. diffusa* group died despite good compliance from the owners. Conservative therapy with enalapril to control hypertension and ampicillin + cloxacillin to prevent urinary infections showed clinical improvement; however, treatment with *B. diffusa* could improve the overall survivability and recovery in CRF dogs.

As *B. diffusa* is a promising alternative treatment modality in CRF, studies addressing the pharmacokinetics of *B. diffusa* extract, especially in renal failure, are necessary



for determining optimum dosage in CRF dogs. Further, including a biopsy examination of kidneys, before and after the therapy, can reveal nephron rejuvenating abilities of the plant, if any.

## CONCLUSION

The beneficial effect of conservative treatment with enalapril to manage CRF in dogs is well-documented.<sup>[41,63-65]</sup> Outcomes with *B. diffusa* root extract treatment were comparable to enalapril. The advantages of *B. diffusa* were faster improvement in most outcome variables like Hb, potassium, phosphorus by day 30 and urinary protein by day 90, and a greater increase in serum potassium in CRF dogs. Also, it must be noted that five CRF dogs in the enalapril group and only two CRF dogs in Punarnava group died between 60 days and 90 days posttreatment. Further, the improvement of several clinical parameters was much earlier in *B. diffusa* root extract treatment.

## REFERENCES

- O'Neill DG, Elliott J, Church DB, McGreevy PD, Thomson PC, Brodbelt DC. Chronic kidney disease in dogs in UK veterinary practices: Prevalence, risk factors, and survival. *J Vet Intern Med* 2013;27:814-21.
- IRIS Canine GN Study Group Standard Therapy Subgroup, Brown S, Elliott J, Francey T, Polzin D, Vaden S. Consensus recommendations for standard therapy of glomerular disease in dogs. *J Vet Intern Med* 2013;27 Suppl 1:S27-43.
- Patel MV, Gupta SN, Patel NG. Effects of Ayurvedic treatment on 100 patients of chronic renal failure (other than diabetic nephropathy). *Ayu* 2011;32:483-6.
- Harini S, Adilaxmamma K, Mohan EM, Srilatha Ch, Raj MA. Antihyperlipidemic activity of chickpea sprouts supplementation in ovariectomy-induced dyslipidemia in rats. *J Ayurveda Integ Med* 2015;6:104-10.
- Venkata Rao KV, Adilaxmamma K, Prasad PE, Alpha Raj M. Hypoglycaemic and hypolipidemic effects of *Cassia auriculata* Linn seed extract in alloxan induced diabetes mellitus. *J Vet Pharmacol Toxicol* 2013;12:82-6.
- Usha Rani M, Gopala Reddy A, Dilip RG, Alpha Raj M. Oxidative stress due to ochratoxin and T-2 toxin either alone or in combination and evaluation of protective role of *Curcuma longa*, *Zingiber officinale*, toxichek and activated charcoal. *Toxicol Int* 2009;16:63-8.
- Bharathi P, Reddy AG, Reddy AR, Alpharaj M. A study of certain herbs against chlorpyrifos-induced changes in lipid and protein profile in poultry. *Toxicol Int* 2011;18:44-6.
- Pavan Kumar Y, Adilaxmamma K, Venkateswarlu U, Chandrasekhara Rao TS, Alpha Raj M. Protective effect of *Trianthema portulacastrum* on cadmium induced toxicity in rats. *J Vet Pharmacol Toxicol* 2012;11:80-4.
- Devi PR, Adilaxmamma K, Rao GS, Srilatha CH, Raj MA. Safety evaluation of alcoholic extract of *Boswellia ovalifoliolata* stem-bark in rats. *Toxicol Int* 2012;19:115-20.
- Velusami CC, Boddapati SR, Hongasandra Srinivasa S, Richard EJ, Joseph JA, Balasubramanian M, *et al.* Safety evaluation of turmeric polysaccharide extract: Assessment of mutagenicity and acute oral toxicity. *Biomed Res Int* 2013;2013:158348.
- Sairam S, Urooj A. Safety evaluation of *Artocarpus altilis* as pharmaceutical agent in wistar rats. *J Toxicol* 2014;2014:980404.
- Chaitanya Kumar TV, Muralidhar Y, Prasad PE, Prasad TN, Alpha Raj M. Evaluation of therapeutic potential of nanosilver particles synthesised using aloin in experimental murine mastitis model. *IET Nanobiotechnol* 2013;7:78-82.
- Chaitanya Kumar TV, Prasad TN, Adilaxmamma K, Alpha Raj M, Muralidhar Y, Prasad PE. Novel synthesis of nanosilver particles using plant active principle aloin and evaluation of their cytotoxic effect against *Staphylococcus aureus*. *Asian Pac J Trop Dis* 2014;4 Suppl 1:S92-6.
- Rendle A. The Classification of Flowering Plants: Dicotyledons. Vol. 2. London, U.K: Cambridge University Press; 1925.
- Yelne MS, Sharma PC, Dennis TJ. Database on Medicinal Plants Used in Ayurveda. Vol. 1. New Delhi, India: Central Council for Research in Ayurveda and Siddha; 2000.
- Singh A. *Boerhaavia diffusa*: An over-exploited plant of medicinal importance in eastern Uttar Pradesh. *Curr Sci* 2007;93:446.
- Kulkarni YR, Keshav AB, Hari KP, Rajan PR. Evaluation of nephro-protective and anti nephro toxic properties of raktapunarnava roots (*Boerhaavia Diffusa*, L.) gokshur fruits (*Tribulus terrestris*, L.) in drug induced nephrotoxicity. *Int Res J Pharm* 2012;3:329-34.
- Ahmed MD, Dutta BK, Rauf AS. Rotenoids from *Boerhaavia repens*. *Phytochemistry* 1990;29:1709-10.
- Lami N, Kadota S, Tezuka Y, Kikuchi T. Constituents of the roots of *Boerhaavia diffusa* L. II. Structure and stereochemistry of a new rotenoid, boeravinone C2. *Chem Pharm Bull* 1990;38:1558-62.
- Kadota S, Lami N, Tezuka Y, Kikuchi T. Constituents of the roots of *Boerhaavia diffusa* L. I. Examination of sterols and structures of new rotenoids, boeravinones A and B. *Chem Pharm Bull* 1989;37: 3214-20.
- Lami N, Kadota S, Kikuchi T. Constituents of the roots of *Boerhaavia diffusa* L. IV. Isolation and structure determination of boeravinones D, E, and F. *Chem Pharm Bull* 1992;39:1863-5.
- Seth RK, Khanna M, Chaudhary M, Singh S, Sarin JP. Estimation of punarnavosides, a new antifibrinolytic compound from *Boerhaavia diffusa*. *Indian Drugs* 1986;23:583-4.
- Jain GK, Khanna NM. Punarnavoside: A new antifibrinolytic agent from *Boerhaavia diffusa* Linn. *Indian J Chem B* 1989;28:163-6.
- Gupta DR, Ahmed B. A new C-methyl flavone from *Boerhaavia diffusa* linn. *Roots*. *Indian J Chem B* 1984;23:682-4.
- Kokate CK, Purohit AP, Gokhale SB. *Pharmacognosy*. 38<sup>th</sup> ed. Pune: Nirali Prakashan Publishers; 2005. p. 537-8.
- Singh RP, Shukla KP, Pandey BC, Singh RG, Singh RH. Recent approach in clinical and experimental evaluation of diuretic action of punarnava with special reference to nephrotic syndrome. *J Res Educ Indian Med* 1992;11:29-36.
- Pareta SK, Patra KC, Mazumder PM, Sasmal D. Aqueous extract of *Boerhaavia diffusa* root ameliorates ethylene glycol-induced hyperoxaluric oxidative stress and renal injury in rat kidney. *Pharm Biol* 2011;49:1224-33.
- Mishra J. Studies on the effect of indigenous drug *Boerhaavia diffusa* Rom. on kidney regeneration. *Indian J Pharm* 1980;12:59.
- Armbrust LJ, Biller DS, Hoskinson JJ, Meier HT, Lora-Michiels M. The basics of renal ultrasonography. *Vet Med* 2011;96:114.
- Walter PA, Feeney DA, Johnston GR, O'Leary TP. Ultrasonographic evaluation of renal parenchymal diseases in dogs: 32 cases (1981-1986). *J Am Vet Med Assoc* 1987;191:999-1007.
- Lucke VM, Kelly DF, Darke PG, Gaskell CJ. Chronic renal failure in young dogs – possible renal dysplasia. *J Small Anim Pract* 1980;21:169-81.

32. Hoppe A, Swenson L, Jonsson L, Hedhammar A. Progressive nephropathy due to renal dysplasia in shih tzu dogs in Sweden: A clinical pathological and genetic study. *J Small Anim Pract* 2008;31:83-91.
33. McGrooty Y. Diagnosis and management of chronic kidney disease in dogs and cats. *In pract* 2008;30:502-7.
34. Mrudula V, George VT, Balachandran C, Manohar MB. Haematobiochemical, urinalysis and urinary enzyme alterations in canine nephritis. *Indian Vet J* 2005;82:826-9.
35. Rubin SI. Chronic renal failure and its management and nephrolithiasis. *Vet Clin North Am Small Anim Pract* 1997;27:1331-54.
36. Eschbach J, Adamson J. Hematologic consequences of renal failure. In: Brenner B, Rector F, editors. *The Kidney*. 9<sup>th</sup> ed. Philadelphia: WB Saunders Co.; 2012.
37. Cowgill LD. Systemic hypertension. In: Kirk RW, editor. *Current Veterinary Therapy IX*. Philadelphia: WB Saunders; 1986. p. 360-4.
38. Jacob F, Polzin DJ, Osborne CA, Neaton JD, Lekcharoensuk C, Allen TA, *et al.* Association between initial systolic blood pressure and risk of developing a uremic crisis or of dying in dogs with chronic renal failure. *J Am Vet Med Assoc* 2003;222:322-9.
39. Buranakarl C, Ankanaporn K, Thammacharoen S, Trisiroj M, Maleeratmongkol T, Thongchai P, *et al.* Relationships between degree of azotaemia and blood pressure, urinary protein: Creatinine ratio and fractional excretion of electrolytes in dogs with renal azotaemia. *Vet Res Commun* 2007;31:245-57.
40. Tylicki L, Rutkowski P, Renke M, Rutkowski B. Renoprotective effect of small doses of losartan and enalapril in patients with primary glomerulonephritis. Short-term observation. *Am J Nephrol* 2002;22:356-62.
41. Lefebvre HP, Brown SA, Chetboul V, King JN, Pouchelon JL, Toutain PL. Angiotensin-converting enzyme inhibitors in veterinary medicine. *Curr Pharm Des* 2007;13:1347-61.
42. Polzin DJ. 11 guidelines for conservatively treating chronic kidney disease. *Vet Med* 2007;102:788-99.
43. Gaitunde BB, Kulkarni HJ, Nabar SD. Diuretic activity of punarnava (*Boerhaavia diffusa*). *Bull Haffkine Inst* 1974;2:24.
44. Dhar ML, Dhar MM, Dhawan BN, Mehrotra BN, Ray C. Screening of Indian plants for biological activity: I. *Indian J Exp Biol* 1968;6:232-47.
45. Chopra GL. *Angiosperms. Systematics and Life Cycle*. Jalandhar, Punjab: S. Nagin and Co.; 1969. p. 361-5.
46. Brenner BM. Nephron adaptation to renal injury or ablation. *Am J Physiol* 1985;249 (3 Pt 2):F324-37.
47. Srinivasan SR, Rajan TS, Dhanapalan P, Tanikachalam M, Gnanaprakasam V. Evaluation of certain routine laboratory tests in the diagnosis of renal insufficiency in canines. *Indian J Vet Med* 1993;13:58-60.
48. Cowgill LD, James KM, Levy JK, Browne JK, Miller A, Lobingier RT, *et al.* Use of recombinant human erythropoietin for management of anemia in dogs and cats with renal failure. *J Am Vet Med Assoc* 1998;212:521-8.
49. Lucke VM, Kelly DF, Darker PG, Gaskell CJ. Chronic renal failure in young dogs-possible renal dysplasia. *J Small Anim Pract* 2008;38:1156-61.
50. Finco Dr. Kidney functions. In: Kaneko JJ, Harvey JW, Bruss ML, editors. *Clinical Biochemistry of Domestic Animals*. 5<sup>th</sup> ed. Iowa: Academic Press;1997. p. 441-81.
51. Polzin DJ, Carl AO, Jacob F, Sheri R. Chronic renal failure. In: Ettinger SJ, Feldman EC, editors. *Text Book of Veterinary Internal Medicine*. 5<sup>th</sup> ed. Philadelphia: WB Saunders, Co.; 2000. p. 1154.
52. Booth K. A case of juvenile nephropathy in a Newfoundland dog. *Vet Rec* 1990;127:596-7.
53. Cetinkaya R, Odabas AR, Selcuk Y. Anti-proteinuric effects of combination therapy with enalapril and losartan in patients with nephropathy due to type 2 diabetes. *Int J Clin Pract* 2004;58:432-5.
54. Grauer GF. Canine glomerulonephritis: New thoughts on proteinuria and treatment. *J Small Anim Pract* 2005;46:469-78.
55. Grauer GF. Measurement, interpretation, and implications of proteinuria and albuminuria. *Vet Clin North Am Small Anim Pract* 2007;37:283-95, vi.
56. Prause LC, Grauer GF. Association of gastrointestinal hemorrhage with increased blood urea nitrogen and BUN/creatinine ratio in dogs: A literature review and retrospective study. *Vet Clin Pathol* 1998;27:107-11.
57. Dey PC, Nath B, Nayak DC, Mukherjee SK. Clinical assessment of Nephtone for renal disorders in dogs. *Phytomedica* 2004;5:125-8.
58. Furuhamo K, Takayama S, Onodera T. Studies on experimental renal damage in rats. I. Analysis of urinary alkaline phosphatase (author's transl). *Nihon Yakurigaku Zasshi* 1982;79:113-21.
59. Valentovic M, Williams P, Carl J 3<sup>rd</sup>, Rankin GO. Urinary enzyme excretion as a parameter for detection of acute renal damage mediated by N-(3,5-dichlorophenyl) succinimide (NDPS) in Fischer 344 rats. *J Appl Toxicol* 1994;14:281-5.
60. Anjaria J, Parabia M, Bhatt G, Khammar R. *A Glossary of Selected Indigenous Medicinal Plants in India*. Ahmedabad: Sristi Innovations; 2002. p. 16.
61. Rosenfield AT. Ultrasound evaluation of renal parenchymal disease and hydronephrosis. *Urol Radiol* 1982;4:125-33.
62. Nyland TG, Mattoon JS, Wisner ER. Ultrasonography of the urinary tract and adrenal glands. In: Nyland TG, Mattoon JS, editors. *Veterinary Diagnostic Ultrasound*. Philadelphia: WB Saunders, Co.; 1995.
63. Chandler ML, Elwood C, Murphy KF, Gajanayake I, Syme HM. Juvenile nephropathy in 37 boxer dogs. *J Small Anim Pract* 2007;48:690-4.
64. Bywater RJ. Penicillins and cephalosporins. In: Brander GC, Pugh DM, Bywater RJ, Jenkins WL, editors. *The Text Book of Applied Veterinary Pharmacology and Therapeutics*. 5<sup>th</sup> ed. London: Bailliere Tindall; 1991. p. 430.
65. Weller RE, Cullen J, Dagle GE. Hyperparathyroid disorders in dog: Primary, secondary and cancer associated. *J Small Anim Pract* 1985;26:329-41.

**How to cite this article:** Oburai NL, Rao VV, Bonath RB. Comparative clinical evaluation of *Boerhavia diffusa* root extract with standard Enalapril treatment in Canine chronic renal failure. *J Ayurveda Integr Med* 2015;6:150-7.

**Source of Support:** Sri Venkateswara Veterinary University, Tirupati, **Conflict of Interest:** None declared.

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